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 Communication to the editor
 

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 THE ISOLATION AND STRUCTURES  
 OF HYGROLIDIN AMIDE AND  
 DEFUMARYLHYGROLIDIN

Sir:

In a previous paper<sup>1)</sup> we described the isolation and structure of hygrolidin (**1**), an antibiotic produced by *Streptomyces hygroscopicus* D-1166 which is specifically active against *Valsa ceratosperma*, the pathogen of apple canker disease. **1** belongs to a new group of 16-membered macro-lide antibiotics. Recently, additional members of this group such as bafilomycins<sup>2)</sup>, a new macrolide substance<sup>3)</sup> and L-681,110<sup>4)</sup> have been reported. Further screening for minor components of **1** resulted in the isolation of two metabolites named hygrolidin amide (**2**) and defumarylhygrolidin (**3**). In this paper, we wish to describe the isolation, structural elucidation and biological activities of these two minor components.

## Isolation

*S. hygroscopicus* D-1166<sup>1)</sup> was maintained on agar slants consisting of glucose 0.4, malt extract 1.0, yeast extract 0.4 and agar 1.5%, pH 7.3. Fermentation was initiated by transferring a loopful of surface growth to large test tubes (2 × 20

cm) each containing 15 ml of PC-I medium (starch 1.0, Polypepton 1.0, molasses 1.0 and beef extract 1.0%, pH 7.2) and incubated at 30°C for 2 days on a reciprocal shaker.

A 2.0% (v/v) transfer of this culture growth was used to inoculate 500-ml Erlenmeyer flasks each containing 100 ml of N medium with the following composition; starch 2.5, soybean meal 1.5, dry yeast 0.2, (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> 0.2, NaCl 0.5 and CaCl<sub>2</sub> 0.4%, pH 7.0. The fermentation was continued for 72 hours at 28°C on a rotary shaker.

Since the production yields of **2** and **3** were very low, and since the organism produced several antibacterial substances such as azalomycin B, azalomycin F<sub>4a</sub> and geldanamycin, antibiotic isolation was monitored by TLC analysis detecting compounds which showed UV absorption similar to that of **1**. **2** and **3** were isolated from mycelium of *S. hygroscopicus* as summarized in Fig. 1. The production yields of **2** and **3** were 0.07 mg/liter and 0.1 mg/liter, respectively.

 Structures of **2** and **3**

**2**, C<sub>33</sub>H<sub>50</sub>O<sub>10</sub>N, SIMS, (M+Na)<sup>+</sup> *m/z* 712, [α]<sub>D</sub><sup>25</sup> +43.4° (c 0.1, MeOH), UV λ<sub>max</sub><sup>MeOH</sup> 245 nm (ε 14,000), 276 (5,800), IR ν<sub>max</sub><sup>KBr</sup> 3400, 1720, 1685, 1675, 1650, 1615, 1250 cm<sup>-1</sup>, was obtained as rod

Fig. 1. Isolation procedures for hygrolidin amide (**2**) and defumarylhygrolidin (**3**).

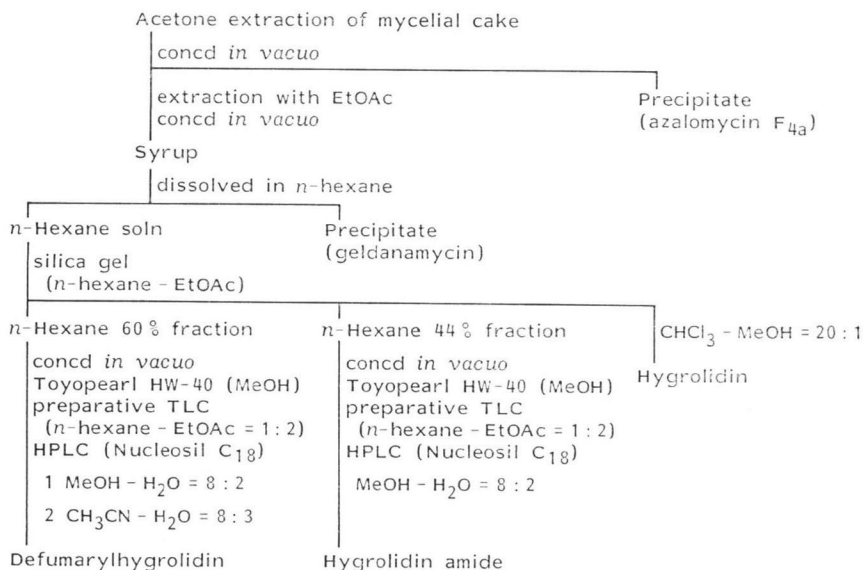


Table 1.  $^{13}\text{C}$  NMR chemical shifts of hygrolidin amide (2) and defumarylhygrolidin (3).

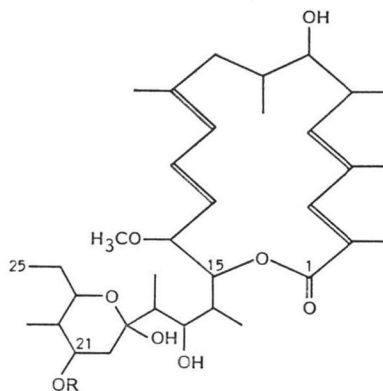
Carbon	2	3	Carbon	2	3
1	171.9	171.9	20	34.4	37.5
2	122.2	122.2	21	73.1	68.0
3	146.4	146.4	22	35.2	38.1
4	134.3	134.3	23	71.3	71.6
5	144.4	144.5	24	25.2	25.3
6	36.7	36.6	25	10.7	10.7
7	81.2	81.1	26	13.7	13.6
8	39.7	39.7	27	15.1	15.1
9	41.2	41.1	28	17.4	17.4
10	142.4	142.5	29	21.5	21.5
11	125.1	125.2	30	20.0	20.0
12	132.5	132.3	31	9.6	9.6
13	127.3	127.3	32	6.9	6.8
14	82.3	82.5	33	4.8	3.9
15	76.0	76.0	34	55.5	55.5
16	37.8	37.8	35	163.9	
17	70.3	70.3	36	132.5	
18	41.6	41.5	37	135.3	
19	99.3	99.2	38	165.0	

Spectra were taken in  $\text{CDCl}_3$  solution and the chemical shifts were expressed in ppm from internal TMS.

crystals which melted at  $138\sim 139^\circ\text{C}$ . The molecular ion peak  $712 (\text{M}+\text{Na})^+$  was shifted to  $728 (\text{M}+\text{K})^+$  when the mass spectrum was taken in the presence of KI.

The  $100\text{ MHz }^{13}\text{C}$  NMR spectrum of **2** is almost completely identical with that of **1**<sup>1)</sup> except for the upfield shift of the carbonyl carbon (C-38) of the fumaric acid residue from  $168.5\text{ ppm}$  to  $165.0\text{ ppm}$  (Table 1). This phenomenon together with the molecular formula of **2** suggested that **2** is an amide derivative of **1**. In agreement with this structure, broad singlet amide protons (area 2H) were observed at  $5.91\text{ ppm}$  in addition to three hydroxyl protons at  $1.62\text{ (d, } J=4.0\text{ Hz)}$ ,  $4.85\text{ (d, } J=4.5\text{ Hz)}$  and  $5.80\text{ ppm (br. s)}$  in the  $400\text{ MHz }^1\text{H}$  NMR spectrum of **2** in  $\text{CDCl}_3$ . When the  $^1\text{H}$  NMR spectrum was taken at  $-10^\circ\text{C}$ , the amide protons changed to two unequal broad singlets at  $6.15$  and  $6.28\text{ ppm}$  and three hydroxyl protons were observed at  $1.85\text{ (d, } J=4.0\text{ Hz)}$ ,  $4.92\text{ (d, } J=4.5\text{ Hz)}$  and  $5.99\text{ ppm (s)}$ . On the other hand, hydroxyl protons could not be observed in the  $^1\text{H}$  NMR spectrum of **1** probably due to rapid exchange of these two protons

Fig. 2. The structures of hygrolidin (1), hygrolidin amide (2) and defumarylhygrolidin (3).



Hygrolidin (1)  $\text{R}=\text{COCH}=\text{CHCOOH}$   
 Hygrolidin amide (2)  $\text{R}=\text{COCH}=\text{CHCONH}_2$   
 Defumarylhygrolidin (3)  $\text{R}=\text{H}$

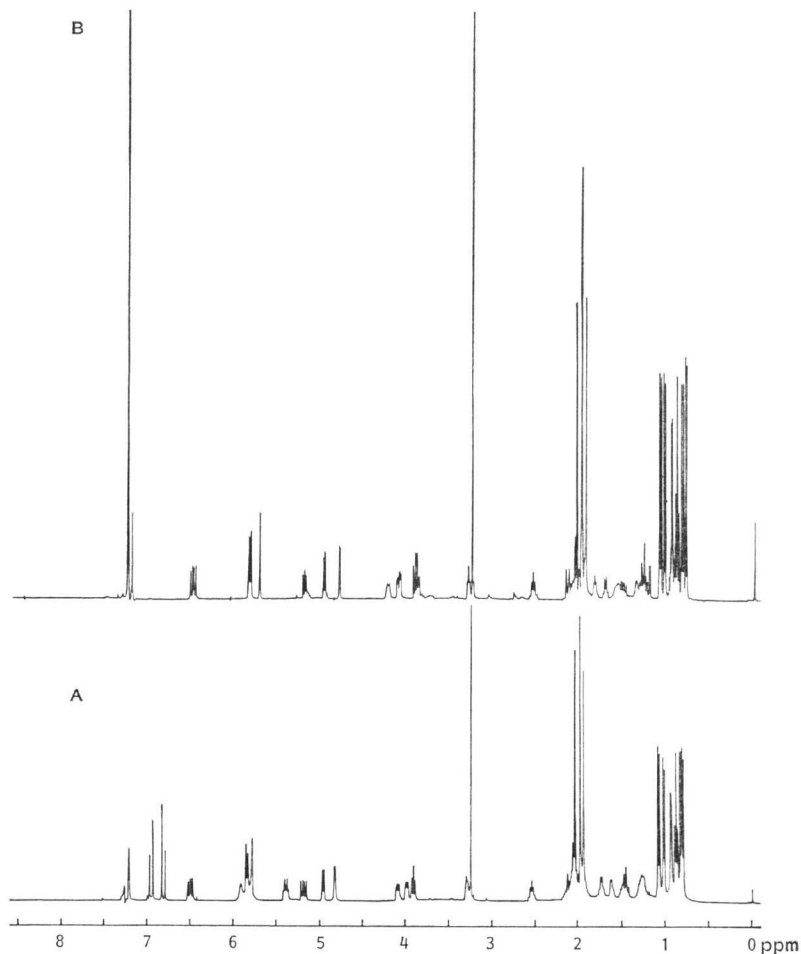
catalyzed by the free carboxylic acid in **1**. Thus, **2** is concluded to be an amide derivative of hygrolidin as shown in Fig. 2.

**3**,  $\text{C}_{34}\text{H}_{56}\text{O}_8$ , SIMS,  $(\text{M}+\text{Na})^+ m/z$  615, was obtained as a white amorphous powder, mp  $95\sim 96.5^\circ\text{C}$ ,  $[\alpha]_D^{25} +49.4^\circ$  ( $c$  0.1, MeOH), UV  $\lambda_{\text{max}}^{\text{MeOH}}$  246 nm ( $\epsilon$  18,400) and 278 (7,500), IR  $\nu_{\text{max}}^{\text{KBr}}$  3400, 1700 (sh), 1670, 1250  $\text{cm}^{-1}$ .

The structure of **3** was determined by detailed analysis of its  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectral data. As shown in Fig. 3, the  $400\text{ MHz }^1\text{H}$  NMR spectrum of **3** is very close to that of **2**. The only difference observed was upfield shift of H-21 from  $5.4\text{ ppm}$  in **2** to  $3.85\text{ ppm}$  in **3** together with the absence of the characteristic AB-type quartet ( $6.82$  and  $6.90\text{ ppm}$ ,  $J=16.0\text{ Hz}$ ) ascribed to the fumaric acid moiety. The  $^{13}\text{C}$  NMR spectral data of **3** (Table 1) were in accord with this structural modification; the resonances due to the fumaric acid moiety disappeared and the C-21 signal showed upfield shift by  $5.1\text{ ppm}$  in the  $^{13}\text{C}$  NMR spectrum of **3**. Thus, **3** is the defumaryl derivative of hygrolidin as shown in Fig. 2 and named as defumarylhygrolidin.

#### Biological Activities

**2** and **3** exhibited selective antifungal activity against *V. ceratosperma* but their activities are much weaker than **1**. The relationship between the concentration and the diameter of inhibition zone is summarized in Table 2. **2** and **3** were inactive against *Bacillus subtilis* and *Escherichia coli* at the concentration of  $100\text{ }\mu\text{g/ml}$ . Cyto-

Fig. 3. 400 MHz  $^1\text{H}$  NMR spectra of hygrolidin amide (2, A) and defumarylhygrolidin (3, B).Table 2. Antifungal activity of hygrolidin amide (2) and defumarylhygrolidin (3) against *V. ceratosperma*.

Concentration ( $\mu\text{g}/\text{ml}$ )	Inhibition zone (diameter, mm)	
	2	3
25.0	20	45
12.5	19	40
6.25	18	37
3.13	17	31.5
1.56	16	27.5
0.78	14	22

Inhibition zone of hygrolidin (1) at 5  $\mu\text{g}/\text{ml}$  was 50 mm.

toxicity of 3 against Friend cell was not observed at 10  $\mu\text{g}/\text{ml}$ , while 2 was toxic at a level of 2  $\mu\text{g}/\text{ml}$  ( $\text{IC}_{50}$ ). Thus, the fumaric acid residue in hygrolidin derivatives seems to play an important role for the biological activities.

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(Received January 31, 1984)

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